



CROTON BONPLANDIANUM IN TRADITIONAL INDIA

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ABSTRACT

In an attempt to strengthen the scientific foundation for the extraction of bioactive components from the plant, the cytotoxic and antioxidant properties of Croton bonplandianum Baill's methanolic and dichloromethane extracts were assessed. Dichloromethane extract was not poisonous, and the methanolic extract had 59.62% DPPH and hydroxyl radical scavenging activity. The reference compound, gallic acid, displayed greater antioxidant activity than the plant extracts. The lethality of both methanolic and dichloromethane extracts on brine shrimp was assessed. An in vitro cytotoxic activity test for methanolic extract yielded an LD50 value of 115.76 (0.0048 - 13.76) µg/ml. The standard medication used was etoposide. The standard medication used was etoposide. Together, these findings support the notion that Croton bonplandianum extracts have significant inherent cytotoxic and antioxidant potentials, which is a strong argument in favor of isolating pure bioactive substances. The plant's parts are widely used in traditional medicine to treat a wide range of conditions, including wound healing, hepatoprotective, body swelling, hypertensive, abdominal dropsy, anti-fungal, anti-microbial, antidiabetic, anti-tumor, and anti-cancer. Because traditional multiplication proceeds slowly, the plant is abundant. Its traditional medical applications have been proven by several in vivo and in vitro tests.

KEYWORDS: *Croton bonplandianus, Antioxidant Activity , anticancer activity medicinal plant, phytochemicals .Pharmacological uses; biological activity; bioactive compounds.*

INTRODUCTION

As a complementary and alternative treatment, herbal medicine has developed to treat a wide range of illnesses brought on by anxiety, industrial dangers, pathogenic microorganisms, stress, and other factors. Even with the availability of contemporary synthetic pharmaceuticals and antibiotics, herbal remedies continue to be used in daily therapy and the medical sector. Many characteristics, including its relative lack of major side effects, affordability, accessibility, and effectiveness, have made herbal therapy a popular and respectable means of treating illnesses even in the present era. Ayurveda, which means "knowledge of long life," started in India during the Vedic era [6]. The two founders of the Ayurvedic medical system, Susrata and Charaka Sanhita, discussed the therapeutic use of several medicinal plants. Ayurvedic literature mentions Croton bonplandianus Baill as one such plant. Therefore, the review is predicated on C. bonplandianus's pharmacological and therapeutic qualities. [5] Diterpenes and phorbol esters, such as 12-orthotrideconeoxy-phorbol-13-acetate (TPA) and myristoyl-phorbol acetate (MPA), are present in Croton bonplandianum seeds. TPA alters prostaglandin metabolism and is a carcinogen [9,10]. Some ethnic groups utilize the plant's fresh juice as a remedy for headaches. Plant latex has the ability to mend cuts and wounds [11,12]. This paper examines various phytochemicals, including tannin, phlobatannin, terpenoid, glycoside, phenolic, flavonoid, steroid, anthraquinone, saponin, alkaloid, cholesterol, carbohydrate, and protein, both qualitatively and quantitatively. The goal is to provide a clear understanding of the phytochemical status of the stem of C. bonplandianus, which will aid future researchers in their pharmacological analysis of this species. Various methods are employed with croton species. According to Salatino et al. (2007), common applications include the management of cancer, constipation, diabetes, dysentery, digestive issues, external wounds, fever, hypercholesterolemia, hypertension, inflammation, intestinal worms, malaria, pain, ulcers, and weight reduction. People in some regions of India used to treat rheumatism by applying a heated poultice made from powdered C. caleornicus leaves (Wilson et al., 1976). Croton nuts, like those from C. megalocarpus, have recently been demonstrated in Kenya to be a more cost-effective source of biofuel than jatropha. In Kenya, the production of one liter of biofuel from Jatropha requires up to 20,000 liters of water, whereas 0.35 liters are produced from wild cotyledon trees. Any region of the body can be affected by a wide range of disorders, collectively referred to as cancer. Neoplasms and malignant tumors are other words that are used. The primary cause of cancer-related mortality is a process known as metastasis. Globally, cancer is the leading cause of mortality (1 in 8). Between 1975 and 2000, the global cancer burden doubled; by 2020, it is expected to double once more, and by 2030, it is predicted to nearly quadruple. Globally, there were about 12 million new instances of cancer and 7 million cancer-related deaths in 2008; by 2030, that number is expected to rise to 20–26 million new cases and 13–17 million deaths [1]. Consequently, it is imperative that cancer be managed, treated, and cured effectively. The potential of alternative medicines may prove beneficial in the management of cancer, which is one of the primary causes



of death globally. At best, conventional medicines just prolong the patient's life through significant side effects. Only add a few years to the patient's life expectancy at most. There is an urgent need for more effective cancer therapies with fewer adverse effects. Therefore, it is necessary to employ alternate theories or methods for cancer prevention [2]. In order to reduce harm to normal tissues and target the various physiological and biochemical processes that promote tumor formation, an integrative approach to cancer patient care should be used. Interestingly, studies in the lab and in clinical trials have shown that herbal remedies may increase the level of efficacy and decrease harmful effects when used in conjunction with chemotherapy. These details made the use of herbal medicine in conjunction with chemotherapy more feasible [3]. Significant advancements in the field of cancer treatment have been accomplished by higher plants. Vinblastine and vincristine, two antileukemic alkaloids derived from the Madagascan periwinkle (*Catharanthus roseus* syn. *Vinca roseus*), are two early examples. A few more anti-cancer medications are homoharringtonine, taxol, and several camptothecin derivatives [4]. One of the common characteristics of cancer is an excess of apoptosis. For multicellular organisms, programmed cell death, or apoptosis, is an essential part of growth and development. Numerous triggers can cause cells to die, and during apoptosis, they do so in a regulated and controlled manner. Damaged cells normally go through apoptosis, but in the case of cancerous cells, changes may have taken place that stop cells from going through this process. Because there is no control over the growth of cells in these situations, the illness may worsen and lead to the development of tumors. Since many cancer treatments involve using radiation or chemicals to harm the cells, mutations in the apoptotic pathway frequently result in cells that are resistant to this kind of attack, making these tumors often tough to eradicate. [7] Many methods can be used to trigger apoptosis in cells grown in vitro. Glucocorticoid exposure to thymocytes is one of the conventional systems. Additional techniques include radiation exposure, topoisomerase inhibitor-treated environments, removing growth factors from growth media, cell cycle disruption, exposure to kinase or phosphatase inhibitors or activators, disruption of Ca²⁺ homeostasis, overexpression of p¹⁹, members of Ced-3/ICE, and many more [8].

Taxonomical Position



Fig. 1. *Croton bonplandianus*^[47]

Kingdom: Plantae
Subkingdom: Tracheobionta
Infrakingdom: Streptophyta
Superdivision: Spermatophyta
Division: Magnoliophyta
Class: Magnoliopsida
Subclass: Rosidae
Order: Malpighiales
Family: Euphorbiaceae
Subfamily: Crotonoideae



Tribe: Crotonaeae

Genus: Croton L.

Species: Croton bonplandianus

Genus Croton

The deciduous shrubs and small trees that make up the Genus Croton can have lepidote scales or stellate hair. Minute requirements apply. Leaf blades have two glands at the base and alternate. Flowers can be either dioecious or monoecious. Racemose inflorescence that is often terminal. Flowers: five merous; males have little disk glands; stamens: five to thirty; females frequently have remnant petals; ovary: three lozenges. Fruit a pill. Small seeds ecaruncle or caruncle.^[15]

Phytochemical Aspect of Croton bonplandianum

The genus Croton contains diverse types of biomolecules. Terpenoids are the predominant secondary metabolite constituents in the genus, chiefly diterpenoids, which may belong to the cembranoid, clerodane, neoclerodane, halimane, isopimarane, kaurane, secokaurane, labdane, phorbol, and trachylobane skeletal types. Triterpenoids, either pentacyclic or steroidal, have frequently been reported from Croton species. Volatile oils containing mono- and sesquiterpenoids, and sometimes also shikimate-derived compounds, are not rare in the genus. Several species have been reported as sources of different classes of alkaloids, a fact that considerably enhances the importance of the genus from a medicinal point of view. Phenolic substances have frequently been reported, among which flavonoids, lignoids, and proanthocyanidins predominate. (Salatino et al., 2007) A crimson sap is produced by a number of Croton species that include proanthocyanidins and/or alkaloids. The latter might be taspine or any of a number of benzyl compounds that resemble oquinolines. Diterpenes, which are related to clerodanes, cembranoids, halimanes, kauranes, labdanes, phorbol esters, trachylobanes, and sarcopetalanes, are highly prevalent in Croton. Certain species have volatile oils, which make them fragrant. From Croton species, representatives of novel chemical classes, such as phenylbutanoids, glycomate alkaloids, and sarcopetalane diterpenes, have been identified. Although laticifers have been reported in Croton species, little anatomical research has been done on the secretory structures of volatile oils yet. There isn't much research on flavonoids using Croton species. The genus has chemical affinities that group species that contain (i) alkaloids, (ii) trachylobanes, and (iii) kauranes and/or labdanes. pharmacological tests have repeatedly confirmed the customary applications of Croton species. The clerodane trans-dehydrocrotonin was the focus of a large portion of pharmacological tests involving Croton compounds (Salatino et al., 2007). 2010 saw the isolation of crotoncaudatin, a novel flavone, along with nine recognized analogues from the stems of Croton bonplandianum Geisel var. tomentosus Hook. Zou et al. (2010) identified the following compounds: 3,5,6,7,8,3',4'-heptamethoxyflavone, tangeretin, nobiletin, 5,6,7,4'-tetramethoxyflavone, sinensetin, kaempferol, tiliroside, kaempferol-3-O-rutinoside, and rutin. Three flavonoids, two diterpenoids, and an indanone derivative are all present in the ethanol extract of Croton steenkampianus leaves (Adeboye et al., 2008). From the methanolic extract of Croton urucuruna, several antimicrobial compounds were isolated, including acetyl aleuritolic acid, stigmasterol, β -sitosterol, campesterol, β -sitosterol-O-glucoside, sonderianin, catechin, and gallicocatechin (Marize et al., 1997). Phytochemically, the plant has been reported to contain rutin (C₁₈ H₃₆ O₁₉) as its main constituent, together with crotosparinine, crotosparine, and its methyl derivative, aphorbol, which play a key role in wound healing (Divya et al., 2011). Apart from this, Croton bonplandianum is a good source of steroids, unsaturated steroids, phenolics, and alkaloids. It also contains flavonols, cardinols, leucoanthocyanins, and flavonoids (Kothale et al., 2011). The plant also contains another two groups of compounds, viz., terpenoids and glycosides, along with flavonoids and alkaloids. The spent residue obtained after biocrude extraction of Croton bonplandianum is rich in biopolymers such as cellulose, hemicellulose, and lignin. This can produce ethanol and oil (Sharma et al., 1990). Jeeshna et al. (2011) investigated the potential of various solvents to extract various groups of compounds from Croton bonplandianum. They found that methanol was significantly more effective than acetone, chloroform, and petroleum ether in extracting alkaloids, flavonoids, glycosides, steroids, phenols, tannins, saponins, and resins (Table 1). Despite the absence of alkaloids and saponins in the chloroform fraction (Jeeshna et al., 2011), According to Ghosh et al. (2013), some identified triterpenoid compounds from the root of C. bonplandianum include sitosterol, ursolic acid, oleanolic acid, and 3-hydroxyurs-12,15-dien of the ursane skeleton. Tiwar et al. (1981) identified an alkaloid from extracts of Croton bonplandianum 3-methoxy-4,6-dihydroxymorphinandien-7-one and norsinoacutine. Alkaloids such as proporphine, isoquinoline dionone, sparsiflorine, crotoflorine, crotosparine, crotosparinine, Nmethylcrotosparine, and N-methylcrotosparinine are found in the plant and its leaves. β -sitosterol, tartaric acid, vomifoliol, uric acid, and tetrahydroglazievine are found in the leaves and stem. Rutin is also present in leaves. (C₁₈ H₃₆ O₁₉). The following compounds were detected in leaves: 16-Hexadecanoyl Hydrazide (88.69%), 1,2-Benzenedicarboxylic Acid, diisooctyl Ester (5.56%), 2-Piperidinone, N-[4-bromo-n-butyl] (2.56%), Phthalic Acid, bis (7-methyloctyl) Ester (1.80%), and Phytol (1.39%). Diterpenes and phorbol esters, such as 12-orthotrideconeoly-phorbol-13-acetat (TPA) and myristoyl phorbol acetate (MPA), are present in Croton bonplandianum seeds. Apart from β -sitosterol, the roots also include Diterpenes and phorbol esters, such as 12-orthotrideconeoly-phorbol-13-acetat (TPA) and myristoyl phorbol acetate (MPA), are present in Croton bonplandianum seeds. The roots include 3-methoxy-4, 6-dihydroxy morphinandien-7-one, and the phenolic quinonoid alkaloid norsinoacutine in addition to β -sitosterol. The hyperaccumulation of copper in this



species is a remarkable discovery. The latex of *C. bonplandianum* contains the phytochemicals 3-methylquinoline (0.44%), 4-methylphenol (6.86%), and mequinol (0.74%). The fruits of *C. bonplandianum* were found to contain fifteen major phytochemicals: 1, propene, 2-nitro-3-(1-cyclooctenyl) (4.58%), 9,12,15-Octadecatrienoic acid, methyl ester (z,z) (41.81%), diazoprogestosterone (19.03%), decanoic acid, ethyl ester (4.86%), and 6,9,12-Octadecatrienoic acid, 13-Tetradecene-11-yn-1-ol (3.47%).^[15]

Analysis of *Croton bonplandianum*

Qualitative Analysis

Chemical tests were carried out on the Chloroform, Methanol, and aqueous, extracts using procedures to identify the phytochemicals as described by Sofowara [14], Trease and Evans [15] and Harborne [16].

Test for Carbohydrates

To 2ml of extract, 1ml of Molisch's reagent and few drops of concentrated sulphuric acid were added. Formation of Purple colour at the inter phase of the two layers indicated the presence of carbohydrates

Test for Amino acids and Proteins

2ml of filtrate was treated with 2-5 drops of ninhydrin solution placed in a boiling water bath for 1-2 minutes and observed for the formation of purple colour.

Test for Tannins

To 1ml of extract, 2ml of 5% ferric chloride was added. Formation of greenish black color indicated the presence of tannins.

Test for Saponins

To 2ml of extract, 2ml of distilled water was added and shaken in a graduated cylinder for 15minutes lengthwise. Formation of 1cm layer of foam indicated the presence of saponins.

Test for Flavonoids

5ml of dilute ammonia solution was added to a portion of the aqueous filtrate of extract followed by addition of concentrated sulphuric acid. Appearance of yellow colouration indicated the presence of flavonoids.

Test for Alkaloids

To 2ml of extract, 2ml of concentrated hydrochloric acid was added. Then few drops of Mayer's reagent were added. Presence of green color indicated the presence of alkaloids.

Test for Quinones

To 1ml of extract, 1ml of concentrated sulphuric acid was added. Formation of red color indicated the presence of quinones

Test for Glycosides

To 1 ml of the extract add few drops of HCl, allowed for 5 minutes for hydrolysis and neutralized with NaOH solution. A few drops of Fehling's solution A and B for few minutes. An orange red precipitate indicates the presence of glycosides.

Test for Terpenoids

To 0.5ml of extract, 2ml of chloroform was added and concentrated sulphuric acid was added carefully. Red brown color formation at the interface indicated the presence of terpenoids.

Test for Phenols

To 1ml of the extract, 2ml of distilled water followed by few drops of 10% ferric chloride was added. Formation of green color indicated the presence of phenols.



Test for Coumarins

To 1 ml of extract, 1ml of 10% Sodium hydroxide was added. Formation of yellow color indicated the presence of coumarins.^[43]

Compound	Water extract	Methanol extract	Chloroform extract	Water extract	Methanol extract	Chloroform extract
Carbohydrates	+	+	+	+	+	+
Amino acids and proteins	+	+	+	+	+	+
Tannins	+	+	+	+	+	+
Saponins	-	-	-	-	-	-
Flavanoids	+	+	+	+	+	+
Alkaloids	-	-	+	-	-	+
Anthrocyanins & β-Cyanins	+	+	+	+	+	+
Quinones	+	+	+	+	+	+
Glycosides	+	+	+	+	+	+
Terpenoids	+	+	-	+	+	-
Phenols	+	+	+	+	+	+
Coumarins	+	+	-	+	+	-

Table 1: Phytochemical screening

RECENT PHARMACOLOGICAL STUDIES

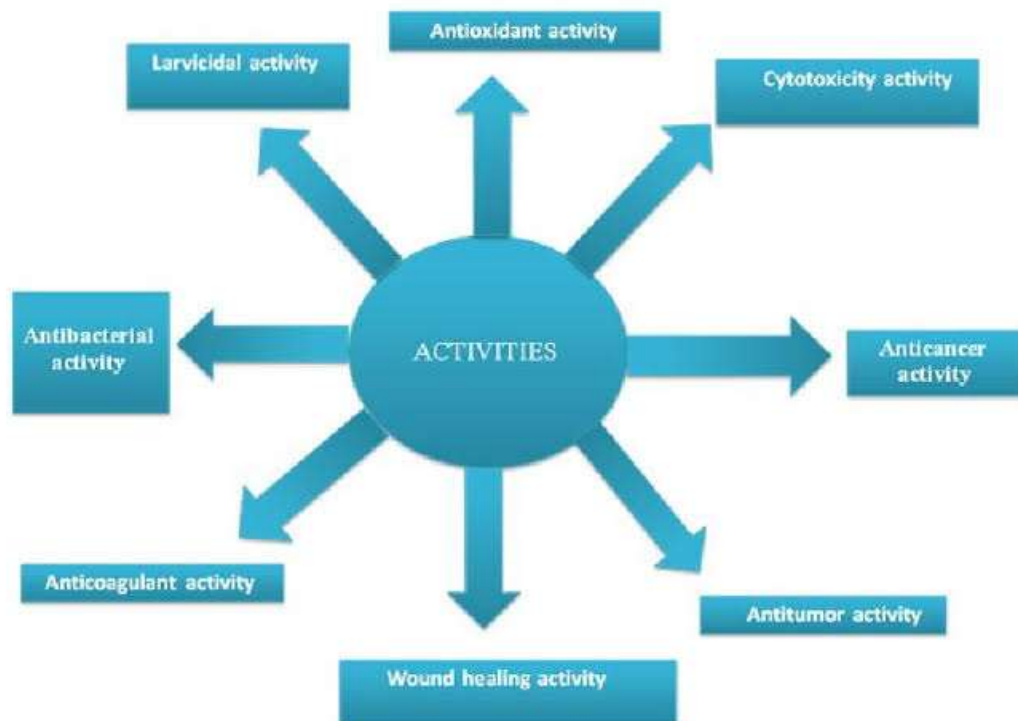


Fig.2 Role of Croton bonplandianum Baill in different activities^[46]



Antioxidant Activity of Croton

In traditional medicine, Croton lechleri sap, often known as "dragon's blood," is applied as a cicatrizant, an anti-inflammatory, and a cancer treatment. When Croton lechleri sap was tested against *Saccharomyces cerevisiae*, a yeast, and against maize plantlets treated with apomorphine and hydrogen peroxide, an oxidative agent, it was discovered that Croton lechleri sap exhibited a notable amount of antioxidant activity against the oxidative damages *Caspomyces cerevisiae* caused. With hydrogen peroxide, on the other hand, the sap's antioxidant activity was only seen in cells that were in the stationary phase of growth. Additionally, the sap was able to shield maize plantlet cells from apomorphine's harmful effects^[30]. A literature survey revealed that the essential oils from northeastern Brazilian Croton species, Croton zenthmeri, Croton nepetaefolius, and Croton argyrophyloides, exhibited good antioxidant activities.^[31] The crude essential oil obtained from the stem bark of Croton urucurana exhibited antioxidant properties. The main components of the antioxidant fraction are α -bisabolol, α -eudesmol, and guaio^[32]. Two of the aromatic acids, vanillic and 4-hydroxybenzoic acid, along with N-methyltyrosine, have been isolated from Croton cajucara. These two aromatic acids have shown remarkable antioxidant activity in other species^[33,34]. Based on such results, C. cajucara could be expected to possess antioxidant properties. Several kaempferol metabolites have proved to be antioxidant agents^[35,36], and C. cajucara leaves also contain two of them, e.g., kaempferol 3,4',7-trimethyl ether and 3,7-dimethyl ether.^[37,38]

Antimicrobial

The agar disc diffusion technique was used to examine the in vitro antibacterial activity of aqueous extracts (say) of Croton bonplandianus against three strains of Gram-positive and four strains of Gram-negative bacteria, according to the National Committee for Clinical Laboratory Standards (1997). Sterile nutrient medium was removed and 100 μ L of bacterial suspension containing 10⁸ colony-forming units (CFU)/mL was mixed with sterile liquid nutritional agar and put to the sterile Petri dishes after the temperature was maintained at 45–50°C. The nutrient-dense agar medium plates were initially dip-soaked in different extract concentrations (10, 25, 50, 75, and As an example, 100 mg/mL. The plates were allowed to harden before filter discs with a diameter of 5 mm were placed on top of them. The plates were incubated at 37 oC for an entire day. The nutritional agar was made by autoclaving it for 15–20 minutes (HI Media Laboratories Limited, Mumbai, India). The diameter of the zone of inhibition—which includes the 5 mm disc diameter—was calculated using a scale. Each experiment was conducted three times to reduce error, and the mean results were presented.^[39,40]

Anticancer Activity

Native to Southeast Asia, Croton tiglium L is a leafy shrub of the Euphorbiaceae family. The plant's seed oil, known as croton oil, or its main active ingredient, 12-Otetradecanoylphorbol-13-acetate (TPA), is an irritant and an inflammatory agent that has been widely used on mice's skin that had previously been exposed to 7,12-dimethylbenz(a)anthracene or other polycyclic aromatic hydrocarbons as a tumor promoter (usual dose: 5–16 nmol, twice a week)^[23,24]. In vitro TPA is an incredibly powerful activator of differentiation in myeloid leukemia cells, even at concentrations 10,000 times lower. TPA has been demonstrated to suppress proliferation, induce apoptosis, or improve differentiation in human tumor cell lines generated from patients suffering from melanoma, prostate, breast, colon, or lung cancer in investigations involving solid tumors^[25,26]. Treatment of LNCaP cells for prostate cancer with clinically feasible TPA doses (1-6 nM) induced apoptosis when these cells were treated with TPA at a concentration that was several times greater, and this treatment led to growth inhibition^[27]. TPA plus ATRA treatment had a synergistic inhibitory effect on the growth of cultured prostate cancer LNCaP cells and also inhibited the growth of existing LNCaP tumors in immunodeficient mice. A combination of TPA and ATRA administered to these tumor-bearing mice caused some tumor regression in all of the treated animals, and tumor regressions were seen in several of the treated mice^[28]. It is unknown what molecular pathways TPA and ATRA use in concert to suppress proliferation and cause apoptosis in LNCaP cells. There has been evidence of a TPA-dependent rise in tumor necrosis factor-alpha (TNF-) in LNCaP cells^[29].

Anthelmintic Activity

The procedure was followed for performing the anthelmintic activity. Due to its morphological and physiological similarities to human intestinal roundworm parasites, the adult Indian earthworm, Pheretima pothuma.^[41] Pheretima pothuma was put in a petridish with three different concentrations of Croton bonplandianum (pet ether, ethanol, and water extract) solutions (20, 40, and 60 mg/ml) in it. Six worms were added to each petri dish, and the worms' paralysis or death was monitored. The mean time for paralysis was recorded when the worm showed no movement at all, with the exception of when it was shaken violently; the time of worm death (min) was recorded once it was determined that the worms did not move when shaken or when the light of outside stimuli. Albendazole was also added as a reference substance in the same way. The test findings were contrasted with samples treated with albendazole (20, 40, and 60 mg/ml) as the reference compound^[42].



Anti-Bacterial Activity

10% w/v test solutions of *C. bonplandianum*'s leaves, fruits, and latex extracts, as well as fresh latex, were made by mixing 500 mg of each extract separately with 5 milliliters of sterile 10% dimethyl sulfoxide (DMSO) in order to examine the antibacterial properties of the plant's materials. To assess fresh latex, extracts from the same source were used to calculate 25, 50, 75, and 100, which had 2.5, 7.5, and 10 mg of antibacterial activity, respectively. At different doses (2.5, 5, 7.5, and 10 mg), the *C. bonplandianum* whole plant and latex extracts were added to each well of Mueller Hinton Agar (MHA) plates that had already been previously infected with the corresponding bacterial cultures. The plates were then incubated at 37 °C for the entire day. In this investigation, 10% DMSO was utilized as the solvent, and the positive and negative control groups were streptomycin (10 µg), respectively. Following incubation, the diameter of the inhibition zone (measured in millimeters) surrounding the well was determined using a zone reader.^[44]

Diphenyl-2-picrylhydrazyl (DPPH) Scavenging Effect

Diphenyl-2-picrylhydrazyl (DPPH) is used to measure the radical scavenging ability of natural products since it can create a stable molecule when it accepts an electron or a hydrogen atom. When an antioxidant and DPPH combine, a free radical known as 1,1-diphenyl-2-picrylhydrazine is produced. The antioxidant drug's scavenging potential is indicated by the degree of decolorization. To 3 ml of extract solution in ethanol at various concentrations (10, 20, 40, 80, and 100 µg/ml), a 0.1 mm solution of DPPH in ethanol was made. One milliliter of this solution was then added. At 517 nm, absorbance was measured after 30 minutes. The reaction mixture's reduced absorbance suggests increased free radical scavenging activity. Using the following formula, the ability to scavenge the DPPH radical was determined. The extract's antioxidant activity was reported as IC. The extract concentration (µg/ml) that suppresses the production of free radicals by 50% is known as the 50% inhibition coefficient (IC value)^[15]

Larvicidal Activity

The study's results, which are shown in the larval susceptibility of *Aegypti* to *C. bonplandianum* methanolic leaf extracts, showed that the methanolic leaf extract was efficient against mosquito larvae. Moreover, it was shown that the effect of larval mortality was dose-dependent. The fourth instar of *A. aegypti* was found to have LC50 and LC90 values of 123.8 ppm and 364.0 ppm, respectively. Therefore, leaves at 124 ppm are recommended for improved vector control. The larvicidal characteristic of *C. bonplandianum* leaf extract could perhaps be attributed to the existence of phorbol derivatives, which are secondary metabolites belonging to the diterpenoids category (Chandel et al., 2005). According to Maria et al. (2006), the essential oils found in four species of a genus, *Croton* are responsible for their larvicidal activity against the mosquito, *A. aegypti*. Nazer et al.^[45]

Traditional Uses

It was discovered that this plant originated in Asia and South America. *C. bonplandianum* is used to treat skin conditions such as ringworm infection, inflammation of the body, and respiratory issues because of its antiseptic qualities. Cholagogue and purgative are the properties of *C. bonplandianum*'s bark and roots^[17,18]. *C. bonplandianum* leaves are used to cure cholera, venereal sores, and cuts and lesions on the body to halt the bleeding. This plant's seeds are used to cure abdominal dropsy, liver problems, acute constipation, and internal abscesses. *C. bonplandianum*'s fresh juice is used to treat headaches^[19,21]. *C. bonplandianum* is widely grown in the rural parts of Malda, West Bengal, and is used as a detergent for fuel. *C. bonplandianum* stems and branches are utilized. Once gathered, the ash is stored for five or six days in a bottle. To make a detergent for cotton clothing, mix ash with warm water. Ethnic people in rural West Bengal, India, use *C. bonplandianum* leaves and roots to treat high fever and snake venom^[20].

CONCLUSION

Traditional methods of treating cancer have involved the widespread usage of numerous medicinal herbs for many generations. One of the biggest genera of flowering plants is *Croton*, and several of its species are widely utilized in ethnomedicine to cure a variety of illnesses, including cancer. As a result, this genus has drawn increasing attention for phytochemical screening and the isolation of any and all anticancer chemicals. The quest for enhanced cytotoxic agents remains a crucial avenue in the advancement of contemporary anticancer pharmaceuticals. The oldest known kind of treatment is phytomedicine. One such plant that is widely utilized in ethnomedical traditions around the globe to cure a variety of ailments is *Croton bonplandianus*. The herb is quite safe and useful for usage as medicine against a variety of ailments, according to traditional and ethnomedical literature. In natural drug discovery, reverse pharmacological methods can be used to explore a plant for a safe and effective medicine.

REFERENCES

1. Mulcahy N. Cancer to Become Leading Cause of Death Worldwide by 2010. *Medscape Medical News*, 2008
2. Reddy L, Odhav B and Bhoola KD. Natural products for cancer prevention: a global perspective. *Pharmacology and Therapeutics*, 2003; 99(1): 1-13.



3. Ruan WJ, Lai M de, Zhou J. Anticancer effects of Chinese herbal medicine, science or myth? *J.Zhejiang Univ Sci B*, 2006; 7(12):1006-1014.
4. Iwu MM, Duncan AR, Okunji CO. In: J. Janick (ed), *Perspective on new crops and new uses*. ASHS Press, Alexandria, VA; 1999; 457-462.
5. Somit Dutta and Tapas Kumar Chaudhuri, *Pharmacological aspect of Croton bonplandianus Bail: A comprehensive review Journal of Pharmacognosy and Phytochemistry* 2018; 7(1): 811-813
6. AYUSH. Department of Ayurveda, Yoga and Naturopathy, Unani, Siddha and Homoeopathy, Ministry of Health and Family Welfare, Govt of India. Available from: <http://www.indianmedicine.nic.in/ayurveda.asp>, 2013.
7. <http://gordonsblog.typepad.com/files/apoptosis.pdf>
8. Saran S. Programmed cell death. *Current Science*, 2000; 78(5): 576
9. Morales A, Perez PP, Mendoza R, Compaqnone R, Suarez AI, Arvelo F. et al. Cytotoxic and proapoptotic activity of ent-16 β -17 α -dihydrokaurane on Human Mammary Carcinoma Cell Line MCF-7. *Cancer Letters*, 2005; 218(131): 109-116
10. Ramesh VM, Hilda A, Manjula VK. Fungitoxic effect of leaf extract of *Croton sparciflorus* on Phytopathic fungi. *Acta Botanica India*. 1995; 23:63-66.
11. Mandal SC, Dewanjee S, Parimala Devi B, Boominathan R, Mazundar R, Mazundar A. Evaluation of antifungal properties of methanolic extracts of some medicinal plants of eastern India, Ninth international congress of the International Society of Ethnobiology. Abstracts for second poster session (15 September 2009) 2004. Available: http://www.kent.ac.uk/sac/ice_2004/posters/wedabsi.html
12. Das AJ, Dutta BK, Sharma GD. *Indian Traditional Knowledge*. 2008; 7(3): 446-454.
13. Chandel KPS, Shukla G, Sharma N. *Biodiversity in medicinal and aromatic plants in India*. National Bureau of Plant Genetic Resources, New Delhi; 1996.
14. Muhammad Naeem Qaisar*, Bashir Ahmad Chaudary, Muhammad Uzair, Sajid Nawaz Hussain Evaluation of Antioxidant and Cytotoxic Capacity of *Croton bonplandianum*. *Baill American Journal of Plant Sciences*, 2013, 4, 1709-1712 <http://dx.doi.org/10.4236/ajps.2013.49208> Published Online September 2013 (<http://www.scirp.org/journal/ajps>)
15. Tanmay Ghosh¹, M. K. Biswas^{2*}, Pradipta Roy¹ and Chiranjib Guin^A Review on Traditional and Pharmacological Uses of *Croton bonplandianum* with Special Reference to Phytochemical Aspect *European Journal of Medicinal Plants* 22(4): 1-10, 2018; Article no.EJMP.40697 ISSN: 2231-0894, NLM ID: 101583475 Published 30th March 2018
16. Aman Singh Patel¹, Asmit Sinha¹, Dr. Pratima Katiyar^{2*} and Dr. Kalpana Kushwaha² Aman Singh Patel¹, Asmit Sinha¹, Dr. Pratima Katiyar^{2*} and Dr. Kalpana Kushwaha² A review of traditional and pharmacology use of *Croton bonplandianum* A *Journal of Pharmaceutical Research* Vol 13, Issue 11, 2024.
17. Maurya SK. Standardization and antioxidant activity of an Ayurvedic formulation "Kushavleha". *international Journal of Green Pharmacy*. 2016; 9(4): 16-26.
18. Chandel KPS, Shukla G, Sharma N. *Biodiversity in medicinal and aromatic plants in India: Conservation and utilization*. New Delhi: National Bureau of Plant Genetic Resources; 1996. p.361
19. Singh NK, Seth A, Maurya SK. *Croton bonplandianum* Baill.: A rich source of essential fatty acids, linoleic and linolenic acid. *Der Pharma Chemica*, 2015, 7(3): 85-88.
20. Ghosh P, Mandal A, Rasul MG. A new bioactive ursane-type triterpenoid from *Croton bonplandianum* Bail. *Journal of Chemical Sciences*. 2013; 125(2): 359-64. 17
21. Reddy KR. Folk medicine from Chittoor District, Andhra Pradesh, India, used in the treatment of jaundice. *International Journal of Crude Drug Research*. 1988; 26(3): 137-40.
22. Runki nath^{1*}, Sarswati roy¹, Blplab de² and M. Dutta choudhury¹ anticancer and antioxidants activity of croton : A REVIEW *Int J Pharm Pharm Sci*, Vol 5, Suppl 2, 63-70
23. Berenblum I. A re-evaluation of the concept of cocarcinogenesis. *Prog. Exp. Tumor Res.* 1969; 11: 21-30
24. Hecker E, Structure-activity relationships in diterpene esters irritant and cocarcinogenic to mouse skin. In: Slaga TJ, Sivak AJ, Boutwell RK, editors. *Mechanisms of Tumor Promotion and Cocarcinogenesis*, Raven New York, 1978; 11-49.
25. Garzotto M, White-Jones M, Jiang Y, Ehleiter D, Liao WC, Haimovitz-Friedman A et al. 12-O-tetradecanoylphorbol-13-acetate induced apoptosis in LNCaP cells is mediated through ceramide synthase. *Cancer Res.* 1998; 58(10) 2260-2264.
26. Rickard KL, Gibson PR, Young GP, Phillips WA. Activation of protein kinase C augments butyrate induced differentiation and turnover in human colonic epithelial cells in vitro. *Carcinogenesis (Lond.)*. 1999; 20(6): 977-984.
27. Konno S, Hsieh TC, Wu JM, Chen Y, Chiao JW, Mallouh C. Growth control of human prostate cancer cells by the phorbol ester TPA: possible involvement of protein kinases. *Anticancer Res.*, 1996; 16(4A): 1843-1849.
28. Zheng X, Chang RL, Cui XX, Avila GE, Lee S, Lu YP. et al. Inhibitory Effect of 12-O-tetradecanoylphorbol-13-acetate Alone or in Combination with All-trans-Retinoic Acid on the Growth of LNCaP Prostate Tumors in Immunodeficient Mice. *Cancer Research*, 2004; 64: 1811-1820.
29. Mizokami A, Gotoh A, Yamada H, Keller ET, Matsumoto T. Tumor necrosis factor- α represses androgen sensitivity in the LNCaP prostate cancer cell line. *J. Urol.*, 2000; 164(3 part 1): 800-805.
30. Lopes e Lopes MI, Saffi J, Echeverrigaray S, Henriques JNP, and Salvador M, Mutagenic and antioxidant activities of *Croton lechleri* sap in biological systems. *Journal of Ethnopharmacology*, 2004; 95(2-3): 437-445.



31. Morais de SM, Catunda Junior FEA, da Silva ARA, Neto JSM, Rondina D, Cardoso JHL. Antioxidant activity of essential oils from Northeastern Brazilian *Croton* species. *Quim.Nova.*, 2006; 29(5): 907-910.
32. Simionatto E, Bonani VFL, Morel AF, Poppi NR, Raposo Junior JL, Stuker CZ, Chemical composition and evaluation of antibacterial and antioxidant activities of the essential oil of *Croton urucurana* Baillon (Euphorbiaceae) stem bark. *J Braz.Chem.Soc.*, 2007 ;18(5):879-885.
33. Hung CY, Yen GC, Antioxidant activity of phenolic compounds isolated from *Mesona procumbens* Hemsl. *J. Agric. Food Chem.*, 2002 50(10) : 299-7.
34. Ohsuqi M, Fan W, Hase K, Xiong Q, Tezuka Y, Komatsu K. et al. Active -oxygen scavenging activity of traditional nourishing tonic herbal medicines and active constituents of *Rhodiola sacra*. *J. Ethnopharmacol.*, 1999 ; 67(1) : 111-9.
35. Marfak A, Trouillas P, Allais DP, Champavier Y, Calliste CA, Duroux JL, Radiolysis of kaempferol in water/methanol mixtures. Evaluation of antioxidant activity of kaempferol and products formed. *J. Agric. Food Chem.*, 2003 ;51(5) : 1270-7.
36. Jonson EL, Schmidt WF, Emche SD, Mossoba MM, Musser SM, Kaempferol (rhmnosyl) glucoside, a new flavonol from *Erythroxylum coca* var. *ipadu*. *Biochem. Syst. Ecol.*, 2003 ; 31 :59-67.
37. Bonina F, Puglia C, Ventura D, Aquino R, Tortora S, Tomaino A. et al. In vitro antioxidant and in vivo photoprotective effects of a lyophilized extract of *Capparis spinosa* L buds. *J. Cosmet. Sci.*, 2002; 53(6): 321-35.
38. Maciel MAM, Pinto AC, Arruda AC, Pamplona SG, Vanderlinde FA, Lapa AJ, et al. Ethnopharmacology, phytochemistry and pharmacology: a successful combination in the study of *Croton cajucara*. *J. Ethnopharmacol.*, 2000; 70(1): 41-55.
39. Khanra K, Panja S, Choudhuri I, Chakraborty A, Bhattacharyya N. Antimicrobial and cytotoxicity effect of silver nanoparticle synthesized by *Croton bonplandianum* Baill. leaves. *Nanomed J.*, 2016; 3(31): 15-2215. doi:10.7508/nmj.2016.01.002
40. CChandra Mohana N, Rakshith D, Ramesha KP, Nuthan BR, Harini BP, Satish S. TLC directed isolation and in silico analysis of antimicrobial metabolite from *Nigrospora sphaerica* inhabiting *Croton bonplandianus* Baill. *South African J Bot.*, 2021; 139: 106-113. doi:10.1016/j.sajb.2021.01.035
41. *Journal of Applied Pharmaceutical Science*, 2012; 02(04): 191-193.
42. Marcocci L, Maguire, Droy-Lefaix JJ. Special reference to that of *Croton Bonplandianum. biloba* extract. *Biophys. Res. Commun*, 1994; 15: 748-755.
43. Satya Prasad M, Suman Joshi DSD, Narendra K, Srinivas K, Srilakshmi Bai J, Lakshmi Chanadana M, Krishna Satya A. Phytochemical And Pharmacological Evaluation Of Euphorbiaceae Family Plant Leaves - *Acalypha Insica* L ., *Croton Bonplandianum* Baill. *Mintage journal of Pharmaceutical & Medical Sciences* | 17-22
44. Vennila V, Udayakumar R. Antibacterial activity of *Croton bonplandianum* (Bail. against some bacterial isolates from infected wounds. *British Microbiology Research Journal*, 2015; 5(1): 83-93.
45. Khanra K, Roy A, Bhattacharyya N. Evaluation of antibacterial activity and cytotoxicity of green synthesized silver nanoparticles using *Hemidesmus indicus* R. Br. *American Journals of Nanoscience and Nanotechnology Research*. 2013;1:1-6.
46. <https://images.app.goo.gl/9QuPTW8fMrW54huv8>
47. <https://images.app.goo.gl/xFiWXR19UCdAzx5m8>