

Volume: 9 | Issue: 11 | November 2024

NOVEL METHOD OF LIPID DRUG MEMBRANE INTERACTION

Ms. Vaishnavi Gadekar¹ , Mr. Laxman Rathod²

¹B Pharmacy 4th year ²M. Pharm 1,2Lokmangal Collage of Pharmacy Wadala

ABSTRACT

A deeper understanding of the molecular mechanisms behind medicine- membrane relations are pivotal for the development of new drugs. To date, a number of biochemical and biophysical styles have been developed to study natural membranes at the molecular position. This review focuses on the advancements and new operations of ultramodern logical ways, including spectrometry, calorimetry, aural seeing, and chromatography, in the study of medicine relations with lipid membranes. The benefits and downsides of these approaches were *compared and precisely considered. also, several biomimetic model membrane types, including liposomes, lipid monolayers, and supported lipid monolayers bilayer, were described. also, a brief overview of the general mechanics underpinning the medicine- membrane commerce process was given.*

KEYWORDS: *Bioanalysis; drug-membrane interactions; drugs; lipid membrane.*

INTRODUCTION

This incredibly complex and varied structure is made up of the main component of the cell membrane, which is a broad spectrum of different lipids, proteins, and polysaccharides. The majority of the continuous lipid bilayer that makes up the cell membrane matrix is composed of amphipathic phospholipids. An essential component of a drug's absorption, distribution, metabolism, and excretion (DME) is the cell membrane, which acts as the cell's border. After being administered, a drug molecule must first enter the bloodstream (the absorption process) and then be delivered to its action regions (the distribution process). Only a few examples of the bio membrane that may be implicated in the action of the medicine are the blood-brain barrier (BBB), the wall of the tiny capillaries lining the stomach, and the barrier membranes in the gastrointestinal system. An increasing amount of research has shown that pharmacological compounds can interact directly or indirectly with the lipid membrane. This interaction may lead to changes in the drug molecules' pharmacological activity, bioavailability, and physicochemical properties, as well as a variety of pharmacological effects and chemical structures. Unfavorable pharmacological interactions with lipid membranes can have negative effects, such as medicine resistance and severe adverse effects. because of poor drug specificity. Anaesthetics, anti-cancer drugs, and non-steroidal anti-inflammatory drugs (NSAIDs) are a few examples. Therefore, understanding the intrinsic interactions between pharmaceuticals and bio membranes is essential for both the pharmaceutical business and biomedical researchers [1].

Analytical techniques for researching drug-membrane interactions

Numerous analytical methods, including spectrometry, calorimetry, chromatography, and acoustical sensing technologies, can be used to study drug-membrane interactions [2].

Chromatographic Techniques

Chromatography is a collection of analytical techniques used to identify, separate, and quantify different substances within a certain range. The foundation of chromatographic techniques is the interaction and differential partition of different substances between a stationary phase and a mobile phase. Technologies like spectroscopy and electrochemical methods are commonly used to enhance the identification of separated components. Depending on the choice of stationary phase and mobile phase, chromatography can be categorized as TLC, liquid chromatography (LC), gas chromatography, capillary liquid chromatography, supercritical fluid chromatography, etc. Here, we will focus on HPLC-based techniques, which are frequently used in studies of drug-membrane interactions [3].

Mechanism of Ion-Association Superior Liquid Chromatography Performance

The HPLCIAM stationary phase was developed to more precisely determine the partitioning of ionic and zwitterionic compounds in various phases. IAM stationary phases, which are marketed by Regis Technologies, are largely composed of covalently attached to porous silica spheres and phospholipid monolayers, mainly phosphatidylcholine. Thus, the application of IAMs may result in a better comprehension of biological activity and partition. Furthermore, compared to the conventional method of determining drug partitioning

SJIF Impact Factor (2024): 8.675| ISI I.F. Value: 1.241| Journal DOI: **10.36713/epra2016 ISSN: 2455-7838(Online)**

EPRA International Journal of Research and Development (IJRD)
Volume: 9 | Issue: 11 | November 2024
Peer Reviewed Journal

Volume: 9 | Issue: 11 | November 2024

in liposome/water systems, AM-HPLC measurement is more suitable for early drug discovery because it is simple, quick, and repeatable [4].Several comparisons have been made between the standard logPandlogDliposome/water and n-octane/water partitioning systems and the lipophilicity as determined by IAM and the capacity factor, log... The results show that AM-HPLC is a more accurate and effective way to determine drug-membrane partition [5].

Liposome Chromatography Paralyzed

According to ILC is an fresh biomimetic system- grounded HPLC [6]. Liposomes are used sterically to screen and dissect passable accoutrements in ILC. The stationary phase is made up of paralyzed gel globules [7]. As opposed to IAMs, the biophysical parcels of the membrane can be altered by varying its composition. ILC columns' lipid terrain. The ILC- calculated lipophilicity indicator(logKs) and other the connections between approaches vary. A relative analysis indicates that only for Significant connections between logKs and structurally analogous substances were set up. IAM, n- octanol/ water, and liposome/ H2O systems all yielded lipophilicity indicators [8]. Only between logKs were significant connections observed in a relative disquisition. and the IAM- deduced lipophilicity indicators, n- octanol/ water,

Chromatography by Electro Kinetics

The capillary electromigration fashion known as electro kinetic capillary chromatography, or electro kinetic chromatography (EKC), is grounded on a combination of HPLC and electrophoresis. EKC measures the analytes' discriminational and electrophoretic mobility. separation of an encircling waterless phase from a lipid dissipation (pseudo-stationary phase) mobile phase) buffer result [9].

Electrochromatography in Capillaries

Capillary electrochromatography (CEC), a lately developed system of capillary liquid chromatography, uses electrostatic inflow to move the mobile phase through a capillary. In CEC measures, more stable lipid coatings are generated due to an IAM stationary phase. Is contained within a capillary made of fused silicon [10]. Demonstrated that direct correlations were. Sixteen structurally different composites in a study comparing CEC and HPLC results [11]. They also refocused out that CEC demanded lower than AM- HPLC. Indeed, with further advanced operation, analyte, fluent, and stationary phase. A recent review looked at the operation of several capillary electrofiltration ways to look at the connections between analytes and lipid membranes [9,12].

Styles of Spectroscopy

An overview of the spectroscopic ways generally used in the study of medicine- membrane relations will be given in this section. These ways includeX-ray diffraction (XRD), luminescence spectroscopy, electron paramagnetic resonance (EPR), vibrational spectroscopy, mass spectroscopy(MS), and small- angle neutron scattering(SANS)[3,13].

Luminescence Spectroscopy

To maintain an eye on intermolecular commerce, luminescence spectroscopy analyzes oscillations in luminescence intensity. luminescence spectroscopy offers remarkable inflexibility, high spatial resolution (down to the position of hundreds of nanometers), and perceptivity (down to the single- patch position) in comparison to contending ways [14]. Since natural luminescence is uncommon in natural systems, fluorescent examinations are generally used. handed a preface to natural membrane luminescence probing [15].

NMR Spectroscopy

NMR is the term for the glamorous parcels of a snippet's nexus. Several of these capitals are set up in lipid motes, similar as 1H, 13C, 31P, 17O, and14N. The introductory idea behind NMR spectroscopy is that some tittles' capitals have a glamorous moment, which causes them to parade distinct energy situations and resonance frequentness when exposed to an external glamorous field [16]. Lipids can also be chemically tagged with other capitals of interest, similar as fluorine(19F) or deuterium(2H) [17].

EPR Spectroscopy

The fashion known as electron spin resonance spectroscopy, or EPR, allows for the direct identification of paramagnetic realities with unmatched electrons [18].

Vibration Spectroscopy

Vibrational spectroscopy uses minimum disturbance to dissect the nuclear vibration parcels of a snippet. Infrared immersion and Raman scattering is its main enterprises. It furnishing the most accurate means of distinguishing between membrane actions, bilayer assembly, and membrane structure and composition, claim [19]. medicine- membrane relations can be anatomized by measuring the medicineconvinced vibrational changes attributable to the specific chemical functional groups inside membrane systems. Fourier transfigure infrared spectroscopy (FTIR) is the most extensively employed infrared spectroscopy fashion for biophysical study. exercising FTIR to

SJIF Impact Factor (2024): 8.675| ISI I.F. Value: 1.241| Journal DOI: **10.36713/epra2016 ISSN: 2455-7838(Online) EPRA International Journal of Research and Development (IJRD)**
Feer Reviewed Journal

Volume: 9 | Issue: 11 | November 2024

track frequency oscillations in the PO2 – str, C=O stretching, and CH2 stretching modes can help us fully understand analyte relations with lipid membranes at the molecular position [20].

X-ray Diffraction

Some X-ray beams will scatter when an entering beam has a wavelength equivalent to the interatomic distances in the sample, demonstrating that XRD measurements are effective [21]. By analyzing the angular distribution of the scattered intensity, X-ray diffraction (XRD) offers a straightforward and non-invasive way to ascertain the sample's structural properties, chemical composition, and physical attributes. Additionally, under near-native conditions, X-ray diffraction (XRD) has the advantage of determining the bilayer thickness of unsupported lipid membranes down to Angstrom length scales [22].

Scattering Neutrons at Small Angles

Both SANS and SAYS are based on similar ideas, with the exception that in SANS, the source of the scattering is the neutron rather than the electron. While SAYS is only sensitive to the hydrophilic region of a lipid bilayer, SANS provides useful information on the hydrophobic tail portion. Therefore, SAYS and SANS may be used as complementary techniques for a detailed structural description of the biological membranes [23].

Colorimetric Methods

The foundation of colorimetric techniques is the evaluation of heat effects associated with drug-membrane interactions [24]. The amount of material involved in the reaction and the heat product's pace are typically associated with the quantity of heat generated or consumed in a chemical reaction. Numerous advanced colorimetric techniques have been used in pharmacology. Pressure perturbation calorimetry (PPC), isothermal ration calorimetry (ITC), and DSC are the most widely used techniques for assessing drug interactions with membrane processes [25].

Differential Scanning Calorimetry (DSC)

It is a non-perturbing technology used to determine a material's heat capacity (CP) as a function of temperature and time. Watson and M. J Neill developed it in 1962, and Chapman used it for the first time in the 1960s to investigate the chemotrophic behavior of bio membranes [26]. The basic notion of ITC and Oscar is the same as the calorimetry of isothermal titration, except operating at a constant temperature and incorporating an titration module[27].In the ITC experiment, aliquots of drugs are retreated in small amounts into liposome solution and vice versa. Until the binding achieves saturation, each injection generates a record of the heat flow. The requirements for thermodynamic values for drug-lipid binding may be determined by binding the created isotherm [28].

Calorimetry of Pressure Perturbation Calorimetry (PPC)

A relatively new thermodynamic technique, measures the change in heat (HQ) that happens when the pressure (UP) above a solution containing proteins or other biomolecules changes [29].

CONCLUSION

The drug's orientation, conformation, and localization within the membrane; the structural stability and phase behavior of the druginserted membrane; the drug's dynamics of interaction with the lipid membrane; and the impact of the drug-membrane interaction on the drug's ADME features are all included. The van der Waals force, hydrogen bonds, and hydrophobic and electrostatic interactions between certain lipid moieties, drug molecules, and membrane proteins are only a few of the variables that can actually affect drugmembrane interactions. Therefore, it is strongly encouraged to use additional analytical techniques in order to have a complete understanding of drug-membrane interaction events. Furthermore, the creation of powerful, novel combinations of techniques, like labon-a-chip hyphenation with MS approaches, would greatly improve the effectiveness of on-site screening in the early stages of drug development.

REFERENCE

- *1. Pereira-Leite,C.;Nunes,C.;Lima,J.L.F.C.;Reis,S.;Lúcio,M.InteractionofCelecoxibwith Membranes:TheRoleofMembraneBiophysicsonitsTherapeuticandToxicEffects.J.Phys.Chem.B2012,116,13608– 13617.10.1021/jp304037vSearchinGoogleScholarPubMed*
- *2. Pignatello,R.DrugBiomembraneInteractionStudies.Sawston,Cambridge:WoodheadPublishing,2013.10.1533/9781908818348SearchinGoog leScholar*
- *3. HewenLi, TaoZhaoEMAILlogoandZhihuaSunEMAILlogFromthejournalReviewsin Analytical Chemistry [https://doi.org/10.1515/revac-](https://doi.org/10.1515/revac-2017-0012)[2017-0012](https://doi.org/10.1515/revac-2017-0012)*
- *4. Yang,C.Y.;Cai,S.J.;Liu,H.;Pidgeon,C.ImmobilizedArtificialMembranes–ScreensforDrug*

SJIF Impact Factor (2024): 8.675| ISI I.F. Value: 1.241| Journal DOI: **10.36713/epra2016 ISSN: 2455-7838(Online) EPRA International Journal of Research and Development (IJRD) Volume: 9 | Issue: 11 | November 2024 - Peer Reviewed Journal**

MembraneInteractions.Adv.DrugDel.Rev.1997,23,229– 256.10.1016/S0169-409X(96)00438-3SearchinGoogleScholar 5. Rutkowska,E.;Pajak,K.;Joźwiak,K.Lipophilicity–MethodsofDeterminationanditsRolein MedicinalChemistry.ActaPol.Pharm.2013,70,3.SearchinGoogleScholar 6. Lundahl,P.;Beigi,F.ImmobilizedLiposomeChromatographyofDrugsforModelAnalysisofDrug-MembraneInteractions.Adv.DrugDel.Rev.1997,23,221– 227.10.1016/S0169-409X(96)00437-1SearchinGoogleScholar 7. Junk,M.J.N.AssessingtheFunctionalStructureofMolecularTransportersbyEPR Spectroscopy;SpringerBerlinHeidelberg:Berlin,Heidelberg, 2012.10.1007/978-3-642-25135-1SearchinGoogleScholar 8. Liu,X.;Fan,P.;Chen,M.;Hefesha,H.;Scriba,G.K.E.;Gabel,D.;Fahr,A.Drug-Membrane InteractiononImmobilizedLiposomeChromatographyComparedtoImmobilizedArtificialMembrane(IAM),Liposome/Water,andOctan-1 ol/WaterSystems.Helv.Chim.Acta2010,93, 203–211.10.1002/hlca.200900233SearchinGoogleScholar 9. Wiedmer,S.K.;Lokajova,J.CapillaryElectromigrationTechniquesforStudyingInteractions betweenAnalytesandLipidDispersions.J.Sep.Sci.2013,36,37– 51.10.1002/jssc.201200829SearchinGoogleScholarPubMed 10. Deeb,S.E.;Wätzig,H.;Elhady,D.A.;Albishri,H.M.;deGriend,C.S.;Scriba,G.K.Recent AdvancesinCapillaryElectrophoreticMigrationTechniquesforPharmaceuticalAnalysis. Electrophoresis2014,35,170.10.1002/elps.201300411SearchinGoogleScholarPubMed 11. Barbato,F.;Grumetto,L.;Carpentiero,C.;Rocco,A.;Fanali,S.Capillary ElectrochromatographyasaNewTooltoAssessDrugAffinityforMembranePhospholipids.J.Pharm.Biomed.Anal.2011,54,893– 899.10.1016/j.jpba.2010.11.037SearchinGoogle Scholar PubMed 12. Wiedmer,S.K.;Lokajova,J.CapillaryElectromigrationTechniquesforStudyingInteractions betweenAnalytesandLipidDispersions.J.Sep.Sci.2013,36,37– 51.10.1002/jssc.201200829SearchinGoogleScholarPubMed 13. HewenLi, TaoZhaoEMAILlogoandZhihuaSunEMAILlogFromthejournalReviewsin Analytical Chemistry https://doi.org/10.1515/revac-2017-0012 14. Weiss,S.FluorescenceSpectroscopyofSingleBiomolecules.Science1999,283,1676– 1683.10.1126/science.283.5408.1676SearchinGoogleScholarPubMed 15. Demchenko,A.P.;Duportail,G.;Oncul,S.;Klymchenko,A.S.;Mely,Y.Introductionto FluorescenceProbingofBiologicalMembranes.MethodsMol.Biol.2015,1232,19–43.10.1007/978-1-4939-1752 5_3SearchinGoogleScholarPubMed 16. Aubin,Y.;Freedberg,D.I.;Keire,D.A.BiophysicalCharacterizationofProteinsinDeveloping Biopharmaceuticals;Elsevier:Amsterdam,2015.SearchinGoogleScholar 17. Osanai,H.;Ikehara,T.;Miyauchi,S.;Shimono,K.;Tamogami,J.;Nara,T.;Kamo,N.AStudyof theInteractionofDrugswithLiposomeswithIsothermalTitrationCalorimetry.J.Biophys.Chem.2013,04,11– 21.10.4236/jbpc.2013.41002SearchinGoogleScholar 18. Junk,M.J.N.AssessingtheFunctionalStructureofMolecularTransportersbyEPR Spectroscopy;SpringerBerlinHeidelberg:Berlin,Heidelberg, 2012.10.1007/978-3-642-25135-1SearchinGoogleScholar 19. Schultz,Z.D.;Levin,I.W.VibrationalSpectroscopyofBiomembranes.Rev.Anal.Chem.2011, 4,343–366.10.1146/annurev-anchem-061010- 114048SearchinGoogleScholar 20. Movasaghi,Z.;Rehman,S.;Rehman,I.U.FourierTransformInfrared(FTIR)Spectroscopyof BiologicalTissues.Appl.Spectrosc.Rev.2008,43,134–179.10.1080/05704920701829043Search in GoogleScholarNasir,M.N.;Benichou,E.;Guez 21. Woolfson,M.M.AnIntroductiontoX-rayCrystallography.Phys.Today1997,50,7074.10.1063/1.882009SearchinGoogleScholar 22. Tyler,A.I.I.;Law,R.V.;Seddon,J.M.MethodsinMembraneLipids;SpringerNewYork:NewYork,NY,2015.SearchinGoogleScholar 23. Cola,E.D.;Grillo,I.;Ristori,S.SmallAngleX-rayandNeutronScattering:PowerfulToolsforStudyingtheStructureofDrug-LoadedLiposomes.Pharmaceutics2016,8, 10.10.3390/pharmaceutics8020010SearchinGoogleScholar 24. Raudino,A.,Sarpietro,M.G.,andPannuzzo,M.,Drug-BiomembraneInteractionStudies.Sawston,Cambridge:WoodheadPublishing,2013.SearchinGoogleScholar 25. Lewis,R.N.A.H.;McElhaney,R.N.EncyclopediaofBiophysics;SpringerBerlinHeidelberg:Berlin,Heidelberg,2013.SearchinGoogleScholar 26. Ladbrooke, B.D.; Williams, R.M.; Chapman, D. StudiesonLecithin-Cholesterol-Water InteractionsbyDifferentialScanningCalorimetryandX-rayDiffraction.Biochim. Biophys. Acta1968,150,333–340.10.1016/0005- 2736(68)90132-6SearchinGoogleScholar 27. Velazquezcampoy,A.;Ohtaka,H.;Nezami,A.;Muzammil,S.;and Freire, Isothermal Titration Calorimetric.Protoc.Stem.CellBiol.2004,62,17.8.117.8.24.10.1002/0471143030.cb1708s23SearchinGoogleScholar 28. Moreno, M.; Garidel, P.; Suwalsky, M.; Howe, J.; Brandenburg, K. The Membrane-Activity of Ibuprofen, Diclofenac, and Naproxen: A Physio-Chemical Study with Lecithin Phospholipids. Biochim. Biophys. Acta2009,1788,1296– 1303.10.1016/j.bbamem.2009.01.016SearchiGoogleScholarPubMedandreferenceno19

29. Heerklotz, H.; Winter, R.; Royer, C.; Seelig, J. Advances in High Pressure Bioscience and Biotechnology, Proceedingsofthe2ndInternationalConferenceonHighPressureBioscienceandBiotechnology,SpringerBerlinHeidelberg: Berlin, Heidelberg, 2003.SearchinGoogleScholar